

KOS Diagnostic Lab (A Unit of KOS Healthcare)



Dr. Vinay Chopra
MD (Pathology & Microbiology)
Chairman & Consultant Pathologist

Dr. Yugam Chopra MD (Pathology) CEO & Consultant Pathologist

NAME : Mrs. RITIKA GULATI

AGE/ GENDER : 37 YRS/FEMALE **PATIENT ID** : 1647408

COLLECTED BY : SURJESH REG. NO./LAB NO. : 012410180048

REFERRED BY : CENTRAL PHOENIX CLUB (AMBALA CANTT) REGISTRATION DATE : 18/Oct/2024 06:22 PM

 BARCODE NO.
 : 01519141
 COLLECTION DATE
 : 18/Oct/2024 06:22PM

 CLIENT CODE.
 : KOS DIAGNOSTIC LAB
 REPORTING DATE
 : 18/Oct/2024 06:31PM

CLIENT ADDRESS: 6349/1, NICHOLSON ROAD, AMBALA CANTT

Test Name Value Unit Biological Reference interval

HAEMATOLOGY COMPLETE BLOOD COUNT (CBC)

RED BLOOD CELLS (RBCS) COUNT AND INDICES

HAEMOGLOBIN (HB)	12.3	gm/dL	12.0 - 16.0
RED BLOOD CELL (RBC) COUNT	4.47	Millions/cmm	3.50 - 5.00
by HYDRO DYNAMIC FOCUSING, ELECTRICAL IMPEDENCE PACKED CELL VOLUME (PCV)	38.7	%	37.0 - 50.0
by CALCULATED BY AUTOMATED HEMATOLOGY ANALYZER MEAN CORPUSCULAR VOLUME (MCV)	86.6	fL	80.0 - 100.0
by CALCULATED BY AUTOMATED HEMATOLOGY ANALYZER MEAN CORPUSCULAR HAEMOGLOBIN (MCH)	27.5	pg	27.0 - 34.0
by CALCULATED BY AUTOMATED HEMATOLOGY ANALYZER MEAN CORPUSCULAR HEMOGLOBIN CONC. (MCHC) by CALCULATED BY AUTOMATED HEMATOLOGY ANALYZER	31.8 ^L	g/dL	32.0 - 36.0
RED CELL DISTRIBUTION WIDTH (RDW-CV) by CALCULATED BY AUTOMATED HEMATOLOGY ANALYZER	13.7	%	11.00 - 16.00
RED CELL DISTRIBUTION WIDTH (RDW-SD) by CALCULATED BY AUTOMATED HEMATOLOGY ANALYZER	44.4	fL	35.0 - 56.0
MENTZERS INDEX by CALCULATED	19.37	RATIO	BETA THALASSEMIA TRAIT: < 13.0 IRON DEFICIENCY ANEMIA: >13.0
GREEN & KING INDEX by CALCULATED	26.53	RATIO	BETA THALASSEMIA TRAIT:<= 65.0 IRON DEFICIENCY ANEMIA: > 65.0
WHITE BLOOD CELLS (WBCS)			
TOTAL LEUCOCYTE COUNT (TLC) by FLOW CYTOMETRY BY SF CUBE & MICROSCOPY	5460	/cmm	4000 - 11000
NUCLEATED RED BLOOD CELLS (NRBCS) by AUTOMATED 6 PART HEMATOLOGY ANALYZER	NIL		0.00 - 20.00
NUCLEATED RED BLOOD CELLS (nRBCS) % by CALCULATED BY AUTOMATED HEMATOLOGY ANALYZER	NIL	%	< 10 %
DIFFERENTIAL LEUCOCYTE COUNT (DLC)			
NEUTROPHILS by Flow cytometry by SF cube & microscopy	79 ^H	%	50 - 70



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Test Name	Value	Unit	Biological Reference interval
LYMPHOCYTES by flow cytometry by Sf cube & microscopy	16 ^L	%	20 - 40
EOSINOPHILS by Flow cytometry by SF cube & microscopy	0_{Γ}	%	1 - 6
MONOCYTES by FLOW CYTOMETRY BY SF CUBE & MICROSCOPY	5	%	2 - 12
BASOPHILS by FLOW CYTOMETRY BY SF CUBE & MICROSCOPY ABSOLUTE LEUKOCYTES (WBC) COUNT	0	%	0 - 1
ABSOLUTE NEUTROPHIL COUNT by FLOW CYTOMETRY BY SF CUBE & MICROSCOPY	4313	/cmm	2000 - 7500
ABSOLUTE LYMPHOCYTE COUNT by FLOW CYTOMETRY BY SF CUBE & MICROSCOPY	874	/cmm	800 - 4900
ABSOLUTE EOSINOPHIL COUNT by Flow cytometry by sf cube & microscopy	0 _L	/cmm	40 - 440
ABSOLUTE MONOCYTE COUNT by flow cytometry by sf cube & microscopy	273	/cmm	80 - 880
ABSOLUTE BASOPHIL COUNT by FLOW CYTOMETRY BY SF CUBE & MICROSCOPY	0	/cmm	0 - 110
ABSOLUTE IMMATURE GRANULOCYTE COUNT by FLOW CYTOMETRY BY SF CUBE & MICROSCOPY PLATELETS AND OTHER PLATELET PREDICTIVE MARKE	55 RS.	/cmm	0.0 - 999.0
PLATELET COUNT (PLT) by HYDRO DYNAMIC FOCUSING, ELECTRICAL IMPEDENCE	322000	/cmm	150000 - 450000
PLATELETCRIT (PCT) by HYDRO DYNAMIC FOCUSING, ELECTRICAL IMPEDENCE	0.29	%	0.10 - 0.36
MEAN PLATELET VOLUME (MPV) by HYDRO DYNAMIC FOCUSING, ELECTRICAL IMPEDENCE	9	fL	6.50 - 12.0
LATELET LARGE CELL COUNT (P-LCC) by HYDRO DYNAMIC FOCUSING, ELECTRICAL IMPEDENCE	61000	/cmm	30000 - 90000
PLATELET LARGE CELL RATIO (P-LCR) by HYDRO DYNAMIC FOCUSING, ELECTRICAL IMPEDENCE	18.9	%	11.0 - 45.0
PLATELET DISTRIBUTION WIDTH (PDW) by HYDRO DYNAMIC FOCUSING, ELECTRICAL IMPEDENCE NOTE: TEST CONDUCTED ON EDTA WHOLE BLOOD	15.7	%	15.0 - 17.0



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Test Name Value Unit Biological Reference interval

IMMUNOPATHOLOGY/SEROLOGY

TYPHOID COMBO SCREEN (TYPHOID ANTIGEN, IgG AND IgM): SERUM

TYPHOID ANTIGEN - SERUM NEGATIVE (-ve) NEGATIVE (-ve)

by ICT (IMMUNOCHROMATOGRAPHY)

TYPHI DOT ANTIBODY IgG NEGATIVE (-ve) NEGATIVE (-ve)

by ICT (IMMUNOCHROMATOGRAPHY)

TYPHI DOT ANTIBODY IgM NEGATIVE (-ve) NEGATIVE (-ve)

by ICT (IMMUNOCHROMATOGRAPHY)

INTERPRETATION:

Typhoid fever is a life threatening illness caused by the bacterium Salmonella typhus. The infection is acquired typically by ingestion. On reaching the gut, the bacilli attach themselves to the epithelial cells of the intestinal villi and penetrate the lamina and submucosa. They are then phagocytosed there by polymorphs and mesenteric lymph nodes, where they multiply and, via the thoracic duct, enter the blood stream. A transient bacteremia follows, during which the bacilli are seeded in the liver, gall bladder, spleen, bone marrow, lymph nodes, and kidneys, where further multiplication takes place. Towards the end of the incubation period, there occurs a massive bacteremia from these sites, heralding the onset of the clinical symptoms.

The diagnosis of typhoid consists of isolation of the bacilli and the demonstration of antibodies. The isolation of the bacilli is very time consuming and antibody detection is not very specific. Other tests include the Widal reaction. The advantage of this test is that it takes only 10-20 minutes and requires only a small amount of stool/serum/plasma to perform. It is the easiest and most specific method for detecting S. typhi infection.

RELATIVE SENSTIVITY OF TYPHOID ANTIGEN DETECTION: 98.7% RELATIVE SPECIFICITY OF TYPHOID ANTIGEN DETECTION: 97.4%

DETECTABLE IGM RESPONSE:

ONSET OF FEVER	PERCENT POSITIVE
4 - 6 DAYS	43.5
6 - 9 DAYS	92.9
> 9 DAYS	99.5

1. This is a solid phase, immunochromatographic ELISA assay that detects specific IgM and IgG Antibodies against the OUTER MEMBRAN PROTEIN(OMP) of the Salmonella species. IgM antibodies appear in the serum 2-3 days post infection and are indicative of a recent infection while the IgG antibodies appear later and are useful for presumptive diagnosis of Enteric fever if the patient presents more than a week after onset of symptoms.

2. This is a useful screening assay for the early detection of Enteric fever and has a high sensitivity. However the test has moderate specificity and false positive results may be obtained in the following situations:

Antibodies against Salmonella may cross react with other antibodies.



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Unrelated infections may lead to production of specific Salmonella antibodies if the patient has previously been exposed to Salmonella infection (ANAMNESTIC RESPONSE).

NOTE:-Rapid blood culture performed during f^t week of infection is highly recommended for confirmation of all IgM positive results. In case the patient has presented after the first week of infection, a thorough clinical correlation and confirmatory Widal test must be performed to establish the diagnosis.



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Test Name Value Unit Biological Reference interval

DENGUE FEVER ANTIGEN NS1 - ELISA (QUANTITATIVE)

DENGUE NS1 ANTIGEN 0.12 INDEX NEGATIVE: < 0.90

QUANTITATIVE BORDERLINE: 0.90 - 1.10

by ELISA (ENZYME LINKED IMMUNOSORBENT ASSAY)

POSITIVE: >=1.10

DENGUE NS1 ANTIGEN NEGATIVE (-ve) NEGATIVE (-ve)
RESULT

by ELISA (ENZYME LINKED IMMUNOSORBENT ASSAY)

INTERPRETATION

DENGUE ANTIGEN NS1					
VALUE	UNIT	RESULT			
< 0.90	INDEX	NEGATIVE (-ve)			
0.90 - 1.10	INDEX	BORDERLINE			
>=1.10	INDEX	POSITIVE (+ve)			

1. The test becomes positive within 0-9 days of exposure to the virus (positive results are obtained within 24 hours of exposure in the overwhelming majority of patients) and generally remains positive till 15 days after exposure. The Dengue NS-1 antigen test is extremely useful in the early diagnosis of the disease thus helping in proper follow up and monitoring of the patients.

2. The IgM antibodies on the other hand take a minimum of 5-10 days in primary infection and 4-5 days in secondary infections to test positive and hence are suitable for the diagnosis of dengue fever only when the fever is approximately one week old.



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MALARIA - P.FALCIPARUM AND P.VIVAX ANTIGEN DETECTION

PLASMODIUM FALCIPARUM ANTIGEN
by ICT (IMMUNOCHROMATOGRAPHY)
DLASMODIUM VIVAY ANTIGEN

NEGATIVE (-ve)

NEGATIVE (-ve)

PLASMODIUM VIVAX ANTIGEN by ICT (IMMUNOCHROMATOGRAPHY)

NEGATIVE (-ve)

NEGATIVE (-ve)

*** End Of Report ***



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