

Dr. Vinay Chopra
 MD (Pathology & Microbiology)
 Chairman & Consultant Pathologist

Dr. Yugam Chopra
 MD (Pathology)
 CEO & Consultant Pathologist

NAME	: Mrs. NISHA	PATIENT ID	: 1753974
AGE/ GENDER	: 32 YRS/FEMALE	REG. NO./LAB NO.	: 012502120009
COLLECTED BY	:	REGISTRATION DATE	: 12/Feb/2025 08:36 AM
REFERRED BY	:	COLLECTION DATE	: 12/Feb/2025 08:37AM
BARCODE NO.	: 01525357	REPORTING DATE	: 12/Feb/2025 11:07AM
CLIENT CODE.	: KOS DIAGNOSTIC LAB		
CLIENT ADDRESS	: 6349/1, NICHOLSON ROAD, AMBALA CANTT		

Test Name	Value	Unit	Biological Reference interval
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CLINICAL CHEMISTRY/BIOCHEMISTRY

GLUCOSE FASTING (F)

GLUCOSE FASTING (F): PLASMA by GLUCOSE OXIDASE - PEROXIDASE (GOD-POD)	138.57^H	mg/dL	NORMAL: < 100.0 PREDIABETIC: 100.0 - 125.0 DIABETIC: > OR = 126.0
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INTERPRETATION

IN ACCORDANCE WITH AMERICAN DIABETES ASSOCIATION GUIDELINES:

1. A fasting plasma glucose level below 100 mg/dl is considered normal.
2. A fasting plasma glucose level between 100 - 125 mg/dl is considered as glucose intolerant or prediabetic. A fasting and post-prandial blood test (after consumption of 75 gms of glucose) is recommended for all such patients.
3. A fasting plasma glucose level of above 125 mg/dl is highly suggestive of diabetic state. A repeat post-prandial is strongly recommended for all such patients. A fasting plasma glucose level in excess of 125 mg/dl on both occasions is confirmatory for diabetic state.




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BARCODE NO.	: 01525357	REPORTING DATE	: 12/Feb/2025 11:46AM
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ENDOCRINOLOGY

ESTRADIOL (E2)

ESTRADIOL (E2): SERUM
 by CMIA (CHEMILUMINESCENT MICROPARTICLE IMMUNOASSAY)

170

pg/mL

FEMALE FOLLICULAR PHASE:
 19.5 - 144.2
 FEMALE MID CYCLE PHASE:
 63.9 - 356.7
 FEMALE PRE OVULATORY
 PHASE: 136.0 - 251.0
 FEMALE LUTEAL PHASE: 55.8 -
 214.2
 POST MENOPAUSAL:< 50.0

INTERPRETATION:

OTHER MATERNAL FACTORS AND PREGNANCY	UNITS	RANGE
Hormonal Contraceptives	pg/mL	15.0 – 95.0
1st Trimester (0 – 12 Weeks)	pg/mL	38.0 – 3175.0
2nd Trimester (13 – 28 Weeks)	pg/mL	678.0 – 16633.0
3rd Trimester (29 – 40 Weeks)	pg/mL	43.0 – 33781.0
Post Menopausal	Pg/mL	< 50.0
MALES:	pg/mL	< 40.0

1. Estrogens are involved in development and maintenance of the female phenotype, germ cell maturation, and pregnancy. They also are important for many other, nongender-specific processes, including growth, nervous system maturation, bone metabolism/remodeling, and endothelial responsiveness.

2. E2 is produced primarily in ovaries and testes by aromatization of testosterone.

3. Small amounts are produced in the adrenal glands and some peripheral tissues, most notably fat. E2 levels in premenopausal women fluctuate during the menstrual cycle.

4. They are lowest during the early follicular phase. E2 levels then rise gradually until 2 to 3 days before ovulation, at which stage they start to increase much more rapidly and peak just before the ovulation-inducing luteinizing hormone (LH)/follicle stimulating hormone (FSH) surge at 5 to 10 times the early follicular levels. This is followed by a modest decline during the ovulatory phase. E2 levels then increase again gradually until the midpoint of the luteal phase and thereafter decline to trough, early follicular levels.

INDICATIONS FOR ASSAY: -

1. Evaluation of hypogonadism and oligo-amenorrhea in females.

2. Assessing ovarian status, including follicle development, for assisted reproduction protocols (eg, in vitro fertilization)

3. In conjunction with luteinizing hormone measurements, monitoring of estrogen replacement therapy in hypogonadal premenopausal women

4. Evaluation of feminization, including gynecomastia, in males.

5. Diagnosis of estrogen-producing neoplasms in males, and, to a lesser degree, females




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6. As part of the diagnosis and work-up of precocious and delayed puberty in females, and, to a lesser degree, males
7. As part of the diagnosis and work-up of suspected disorders of sex steroid metabolism, eg: aromatase deficiency and 17 alpha-hydroxylase deficiency
8. As an adjunct to clinical assessment, imaging studies and bone mineral density measurement in the fracture risk assessment of postmenopausal women, and, to a lesser degree, older men
9. Monitoring low-dose female hormone replacement therapy in post-menopausal women
10. Monitoring antiestrogen therapy (eg, aromatase inhibitor therapy).

CAUSES FOR INCREASED E2 LEVELS:

1. High androgen levels caused by tumors or androgen therapy (medical or sport performance enhancing), with secondary elevations in E1 and E2 due to aromatization
2. Obesity with increased tissue production of E1
3. Decreased E1 and E2 clearance in liver disease
4. Estrogen producing tumors
5. Estrogen Ingestion




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PROGESTERONE

PROGESTERONE: SERUM <i>by CMIA (CHEMILUMINESCENT MICROPARTICLE IMMUNOASSAY)</i>	0.16	ng/mL	MALES: 0.21 - 2.10 NON PREGNANT WOMEN MID FOLLICULAR PHASE: 0.29 - 1.55 MID LUTEAL PHASE: 5.11 - 18.78 PREGNANT WOMEN FIRST TRIMESTER: 4.69 - 51.31 SECOND TRIMESTER: 19.24 - 45.55
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INTERPRETATION:

EXPECTED VALUES OF PROGESTERONE DURING PREGNANCY

	UNITS (ng/mL)
First trimester (0 - 12 Wweeks)	15.8 - 46.0
Second trimester (13 - 28 Wweeks)	15.6 - 74.0
Third trimester (29 - 40 Wweeks)	45.0 - 143.0
Post Menopausal	< 1.40

1. Progesterone is produced by the adrenal glands, corpus luteum, and placenta.
 2. After ovulation, there is a significant rise in serum Progesterone levels as the corpus luteum begins To produce progesterone in increasing amounts. This causes changes in the uterus, preparing it for implantation of a fertilized egg. If implantation occurs, the trophoblast begins to secrete human chorionic gonadotropin, which maintains the corpus luteum and its secretion of progesterone. If there is no implantation, the corpus luteum degenerates and circulating progesterone levels decrease rapidly, reaching follicular phase levels about 4 days before the next menstrual period.

The test is indicated for:

1. Ascertaining whether ovulation occurred in a menstrual cycle
2. Evaluation of placental function in pregnancy
3. Workup of some patients with adrenal or testicular tumors

NOTE:

In patients receiving therapy with high biotin doses (ie, >5 mg/day), no specimen should be drawn until at least 8 hours after the last biotin administration.

*** End Of Report ***




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